



Innate and Adaptive Immune Responses to Adeno-associated viral Gene Therapy in the Severe Hemophilia A Dog Model

P. Batty, A. Mo, A.M. Ismail, B. Yates, A. Gonzalez, B. Handyside, L. Harpell, D. Hurlbut, A. Menard, A. Pender, L. Razon, C.R. Sihn, A. Winterborn, S. Fong, D. Lillicrap

OC 01.5 : Gene Therapy I : Saturday, July 9, 2022



Disclosures for Paul Batty

In compliance with COI policy, ISTH requires the following disclosures to the session audience:

Research Support/P.I.	Octapharma, BioMarin, Grifols
Employee	No relevant conflicts of interest to declare
Consultant	No relevant conflicts of interest to declare
Major Stockholder	No relevant conflicts of interest to declare
Speakers Bureau	No relevant conflicts of interest to declare
Honoraria	Octapharma, BioMarin, Pfizer, Novo Nordisk Institute for Nursing and Medication Education (IMNE)
Scientific Advisory Board	BioMarin

Canine Hemophila A

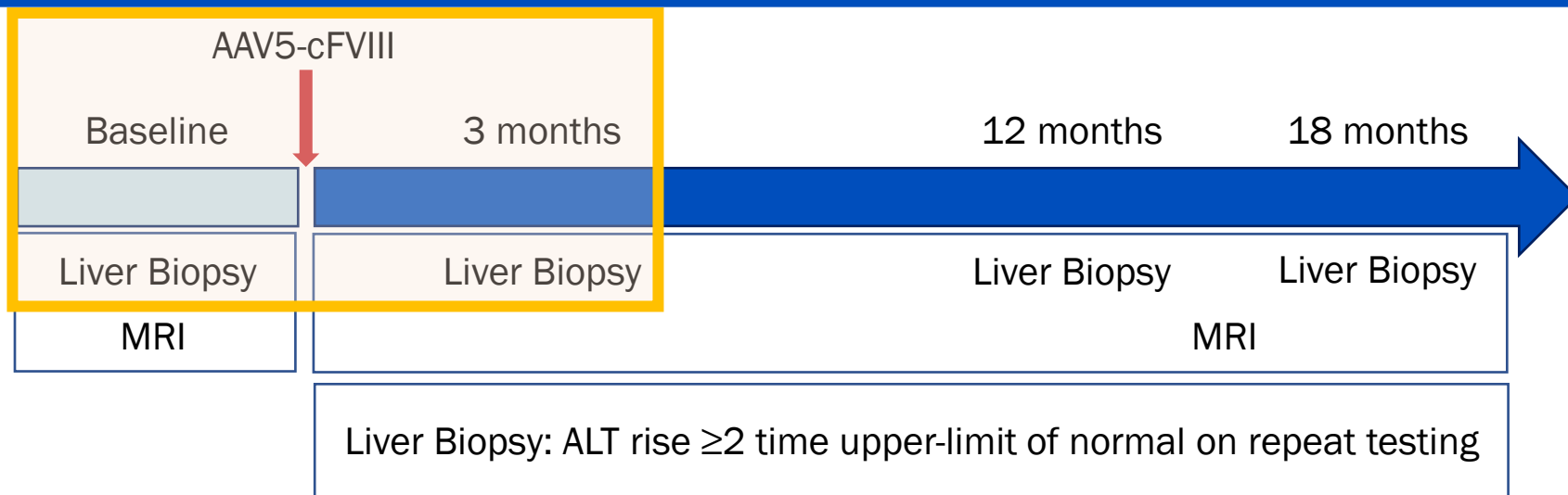


- Similar Genotype
 - Intron-22 inversion like *F8* mutation ¹
- Similar Clinical Phenotype
 - Spontaneous bleeding events
 - Treated with recombinant canine FVIII
 - Inhibitor-prone c. 25% incidence
- Long-term follow-up >10 years ²

1) Hough C et al. Thromb Haemost. 2002; 87(4):659-65.

2) Batty P et al. Blood. 2022; Epub ahead of print. PMID: 35405003.

Study Aims & Outline



- Evaluate safety & efficacy of AAV5-cFVIII in a hemophilia A dog model
- Evaluate factors resulting in variable expression following AAV5-cFVIII
- Evaluate early (<3 months) innate & adaptive immune response to AAV5-cFVIII

Cohort 1 : Non-Codon Optimized AAV5-cFVIII

B-domain deleted canine F8



ITR = Inverted terminal repeat. HLP = Hybrid liver promoter

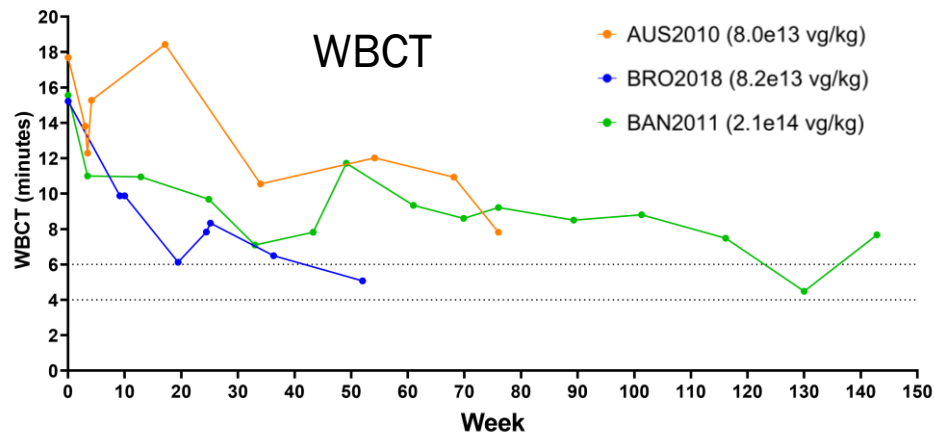
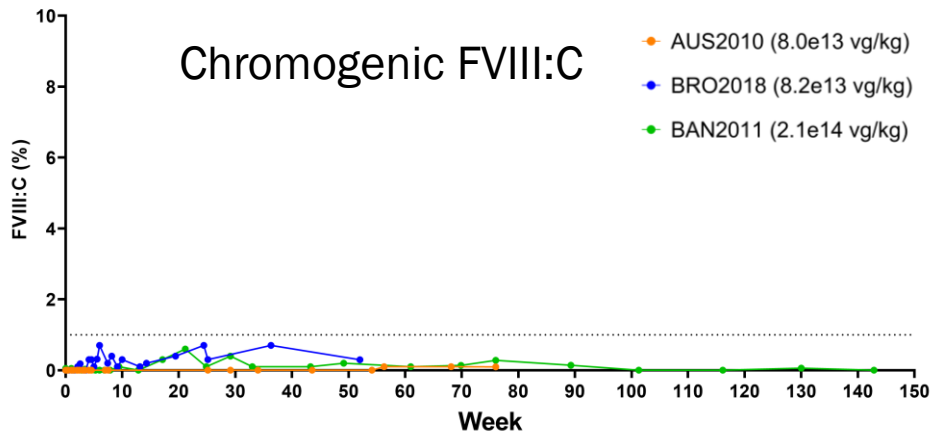
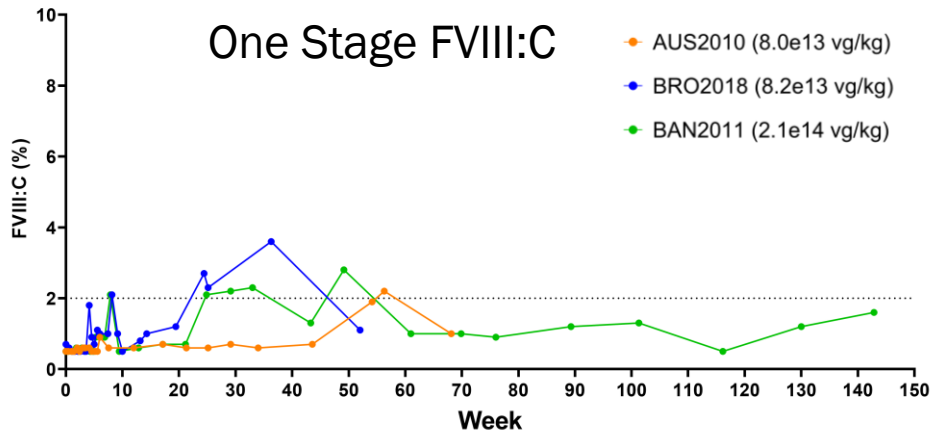
PA = Polyadenylation sequence

Cohort	ID	Dose (vg/kg)	Treatment Age (years)	Follow Up (years)
1	AUS2010	8.0e13	9.0	1.5 #
	BRO2018	8.2e13	0.8	2.8 *
	BAN2011	2.1e14	8.3	1.0 #

Response = Chromogenic FVIII:C >1%

Data capture 29/04/2022. * Follow-up ongoing. # Study end-point reached

No Significant FVIII Expression with AAV5-nco-cFVIII



- Reduction in Whole Blood Clot Time (WBCT) despite no significant FVIII expression
- Similar findings seen in non-responders in our previous study¹

Cohort 2 : Codon Optimized AAV5-cFVIII

B-domain deleted canine F8



ITR = Inverted terminal repeat. HLP = Hybrid liver promoter

PA = Polyadenylation sequence

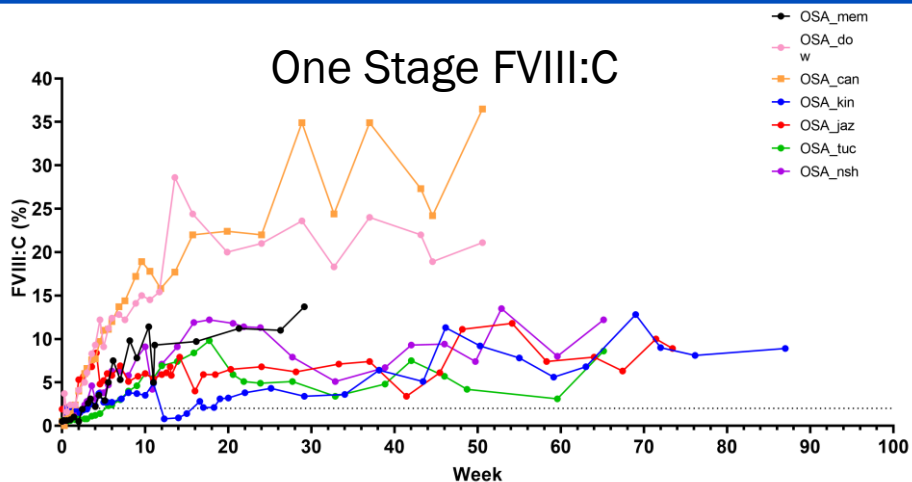
Cohort	ID	Dose (vg/kg)	Treatment Age (years)	Follow Up
2	NAS2018	6.0e13	2.4	1.3 *
	TUC2018	6.0e13	2.4	1.3 *
	KIN2015 (V3)	6.8e13	5.2	1.8 *
	JAZ2013	6.8e13	6.9	1.4 #
3	CAN2017	2.0e14	3.8	1.0 *
	DOW2016	2.0e14	4.9	1.0 *
	MEM2018 (INH)	2.0e14	3.0	0.6 *

Response = Chromogenic FVIII:C >1%. V3 = HLP-co-cFVIII-V3. INH = FVIII inhibitor history

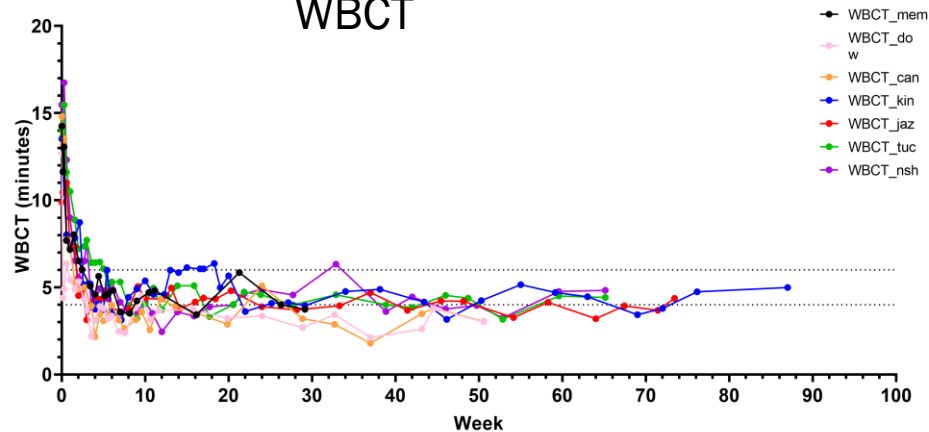
Data capture 29/04/2022. * Follow-up ongoing. # Study end-point reached

Dose Dependent FVIII Expression with AAV5-co-cFVIII

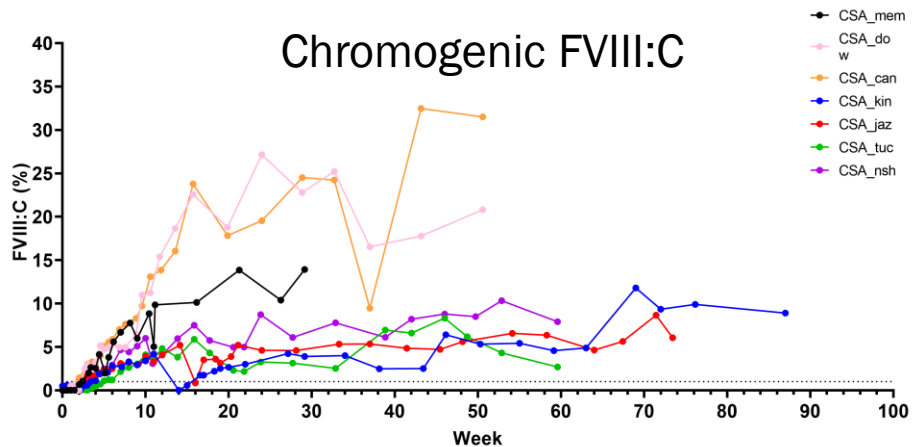
One Stage FVIII:C



WBCT

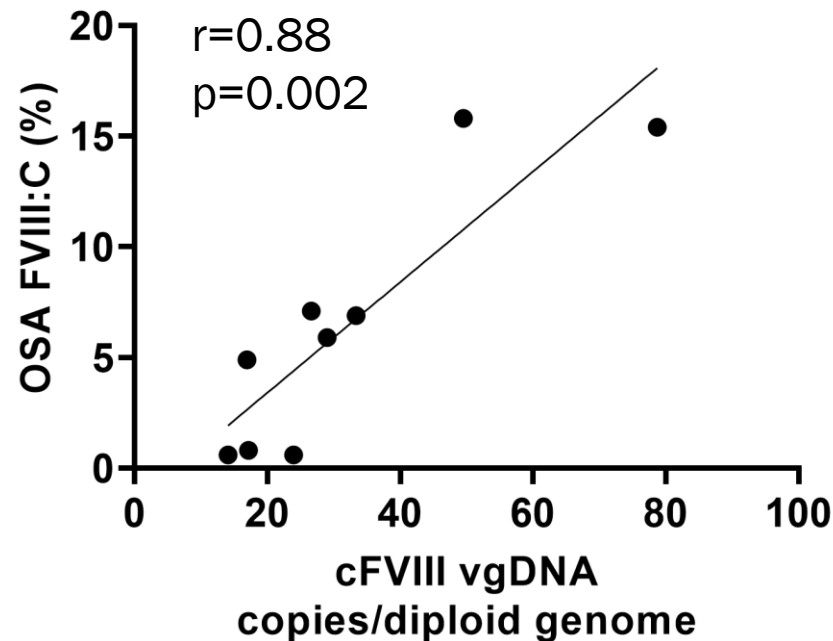
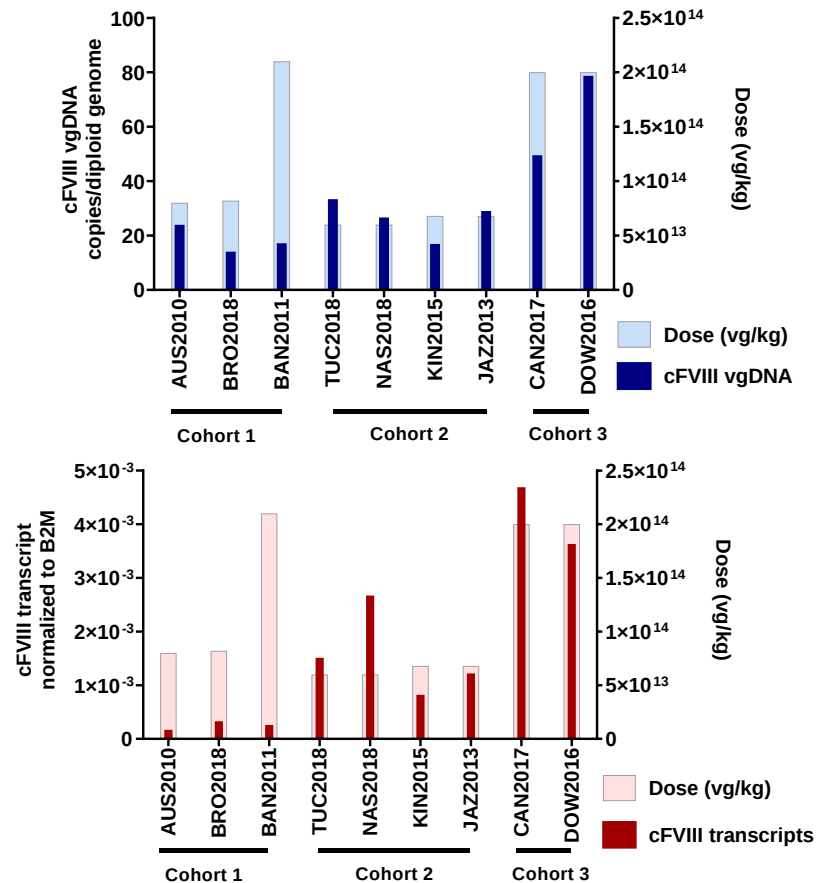


Chromogenic FVIII:C



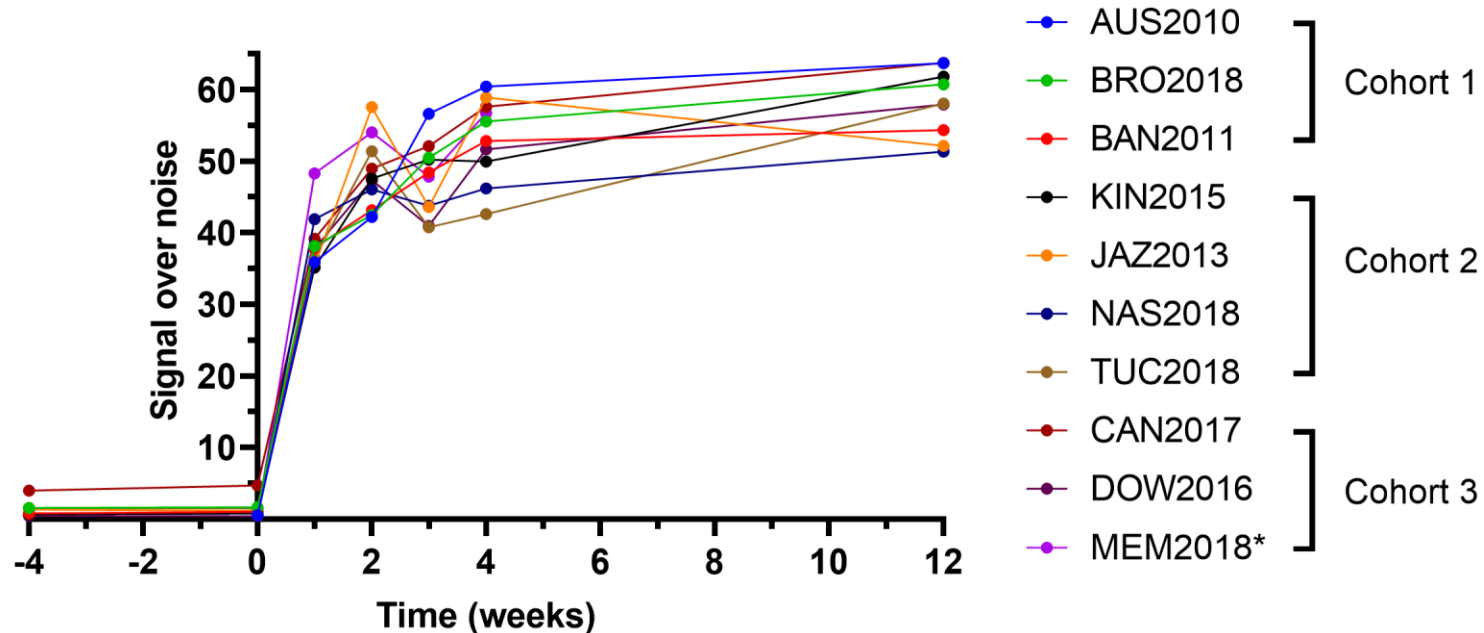
Cohort	ID	Dose (vg/kg)
2	NAS2018	6.0e13
	TUC2018	6.0e13
	KIN2015 (V3)	6.8e13
	JAZ2013	6.8e13
3	CAN2017	2.0e14
	DOW2016	2.0e14
	MEM2018 (INH)	2.0e14

AAV5-cFVIII Detectable in All Dogs with Dose Response and Strong Correlation between Liver Vector Genomes and FVIII Expression



- Strong correlation between liver Vector Genome DNA & FVIII:C at 3 months

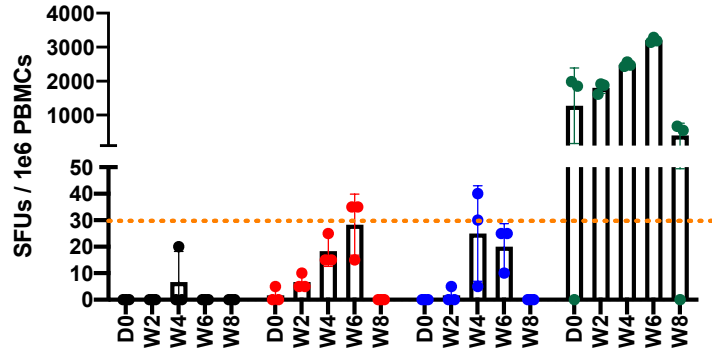
Capsid Antibodies Seen Early Post AAV-cFVIII



- Early capsid antibody response seen at D7 post AAV-cFVIII
- Further studies to assess AAV5 neutralizing antibodies using a cell-based transduction inhibition assay ongoing

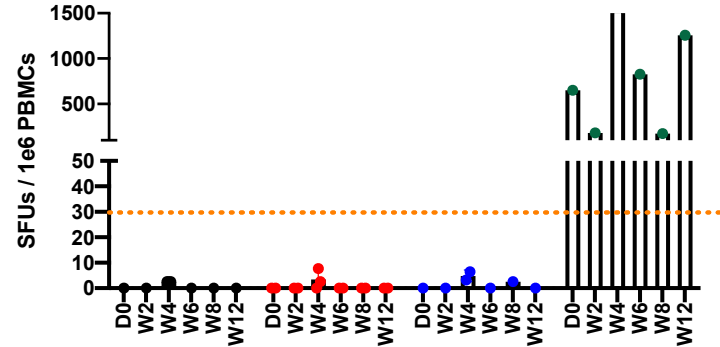
No Significant Cytotoxic T-Cell Response* Observed

BR02018 (Cohort 1, 8.2e13 vg/kg)

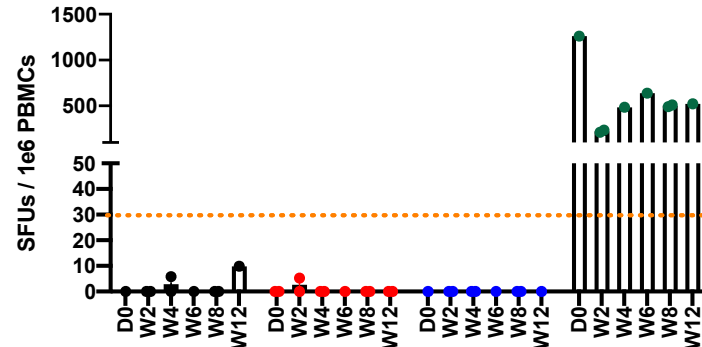


- DMSO
- AAV Peptide Pool 1
- AAV Peptide Pool 2
- Positive Control
- Limit of detection

NAS2018 (Cohort 2, 6e13 vg/kg)

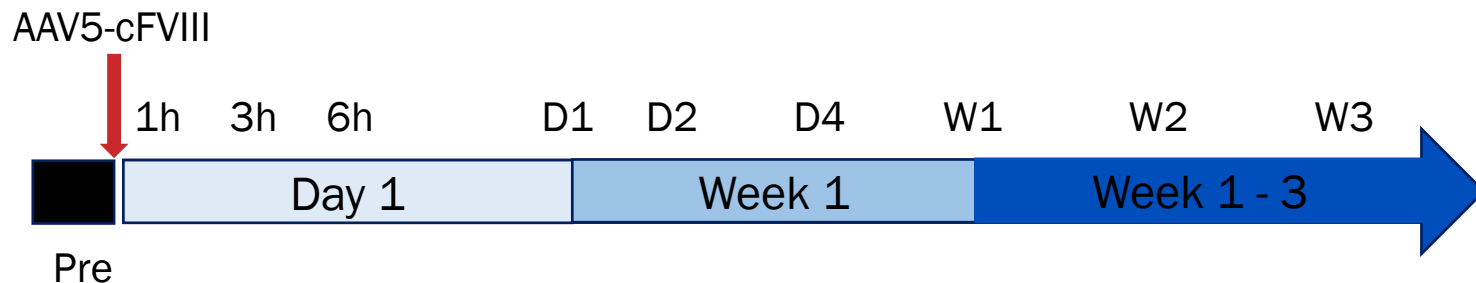


CAN2017 (Cohort 3, 2e14 vg/kg)



* As measured by ELISpot INF γ assay

Evaluation of Early Immune Biomarkers to AAV5-co-FVIII



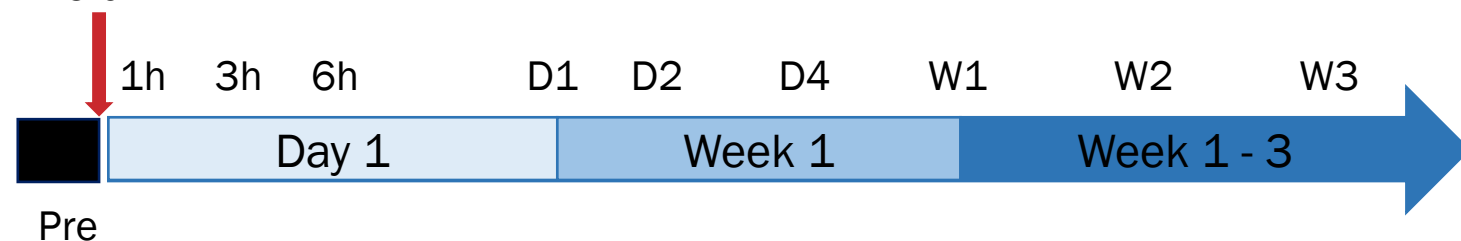
Cohort	ID	Dose (vg/kg)
2	NAS2018	6.0e13
	TUC2018	6.0e13
3	CAN2017	2.0e14
	DOW2016	2.0e14
	MEM2018 #	2.0e14

Biomarkers	
Cytokines ¹	GM-CSF, IFN γ , IL-2, IL-6, IL-7, IL-8, IL-10, IL-15, IL-18, IP-10, KC-like, MCP-1, TNF α , TGF β -1, TGF β -2, TGF β -3
Complement	CH50 (Classical pathway)

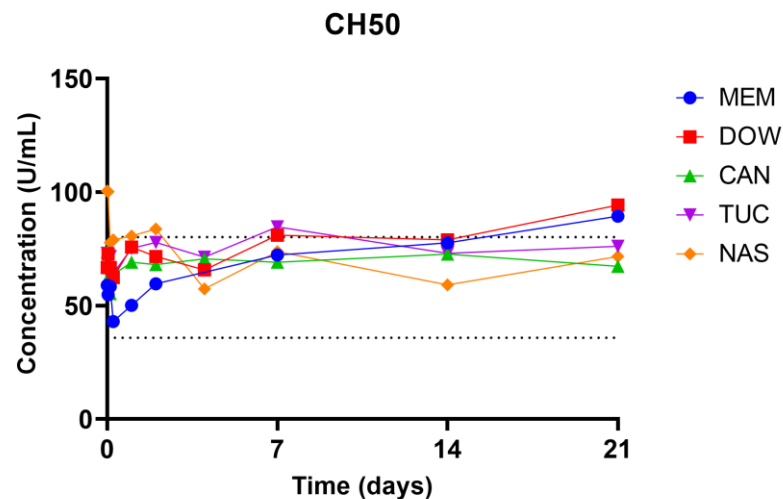
1) Canine Cytokine/Chemokine 13-Plex Discovery Assay® Array (Eve Technologies)

No Complement Activation (CH50) Seen Following AAV5-cFVIII

AAV5-cFVIII

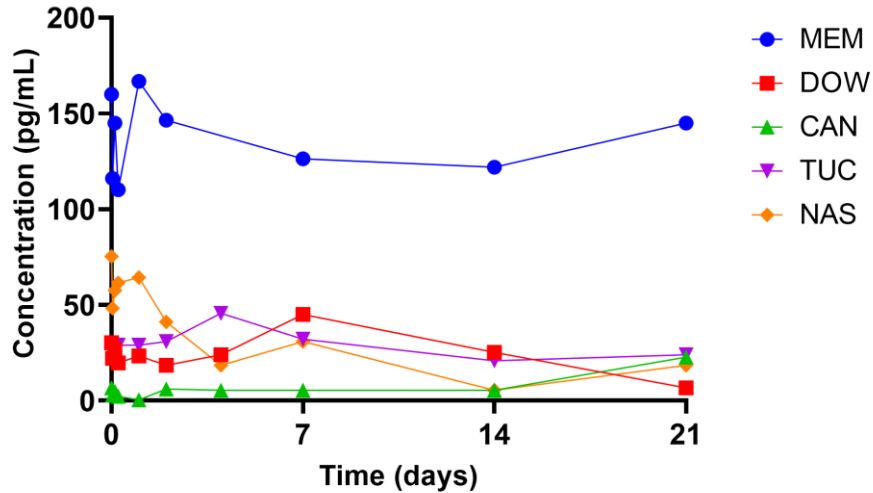


Cohort	ID	Dose (vg/kg)
2	NAS2018	6.0e13
	TUC2018	6.0e13
3	CAN2017	2.0e14
	DOW2016	2.0e14
	MEM2018 #	2.0e14

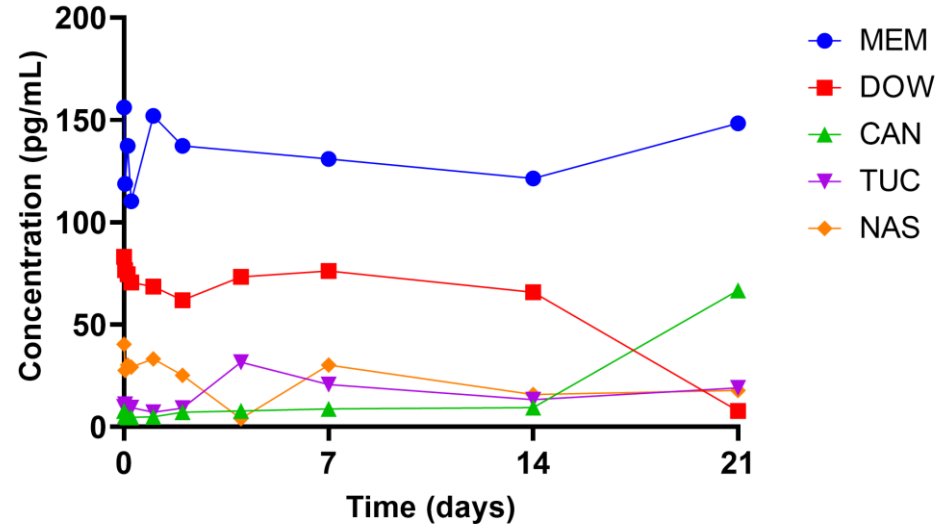


No Significant Elevation in Most Cytokines Analyzed Following AAV5-cFVIII

IL2 D21

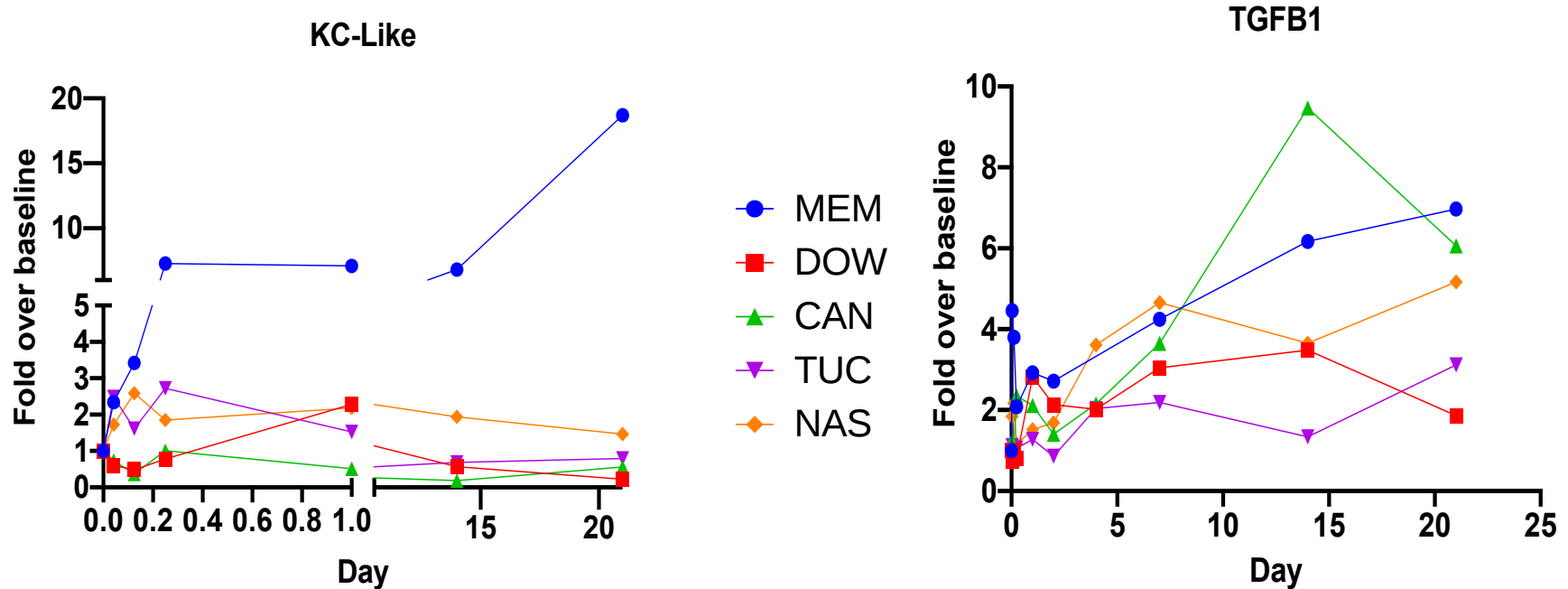


IL6 D21



- Different dogs displayed individual cytokine profiles
- Higher TNF- α , IL2, IL6, IL7, IL15 & IL18 in one dog with an inhibitor history
- No change in most cytokines levels

Minor Elevation of Cytokines Observed Following AAV5-cFVIII



- Minor transient elevation in KC-like within 24 hours seen in 4/5 dogs
- Mild TGFB1 increases with time up to day 21
- No elevation in liver enzymes (ALT, AST or ALP) up to day 21

Summary

- Codon Optimized AAV5-cFVIII resulted in therapeutic cFVIII:C expression with dose response
- Mechanistic studies are ongoing into WBCT improvement without cFVIII:C expression following non-codon optimized AAV5-cFVIII
- AAV5-HLP-cFVIII was detected in the liver in all animals at 3 months, with correlation between VG and cFVIII mRNA copies
- Capsid antibodies seen within 7 days of AAV5-cFVIII
- No significant elevations of innate immune response was detected

Acknowledgments

- Haemophilia colony dogs
- Queen's University
 - Alex Menard
 - Lori Harpell
 - Abbey Pender
 - David Hurlbut
- BioMarin
 - Sylvia Fong
 - Bridget Yates



Contact:

pab7@queensu.ca

[@paul_batty1](https://twitter.com/paul_batty1)