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- Duchenne muscular dystrophy (DMD) is a neuromuscular disorder characterized by progressive muscle weakness that can be caused by frameshift variants that produce truncated and non-functional dystrophin protein^{1,2}
- Nivudirsen (BMN 351) is an antisense oligonucleotide (ASO) designed to exclude exon 51 from the dystrophin pre-mRNA to restore the open reading frame and enable synthesis of functional, near full-length dystrophin protein in exon 51 skip-amenable individuals^{3,4}
- Study 351-201 (NCT06280209) is an ongoing, open-label, phase 1/2 clinical trial of nivudirsen in participants with DMD amenable to exon 51 skipping
- The measurement of exon 51 skipping and dystrophin protein can provide early pharmacodynamic assessments of ASOs under clinical investigation
 - Immunoaffinity ultra-performance liquid chromatography coupled with tandem mass spectrometry (IA-UPLC-MS/MS) offers an improved quantitative measure of dystrophin protein compared with semiquantitative western blot methods

- Three key pharmacodynamic biomarkers were measured in participant muscle biopsy tissue from the 351-201 clinical trial to evaluate early participant responses from the first 2 of 3 total nivudirsen doses

- Tissue homogenate aliquots were used to measure specific mRNA targets by droplet digital PCR (ddPCR), dystrophin, and α -actinin-2 proteins by IA-UPLC-MS/MS, and total protein by the bicinchoninic acid (BCA) assay
- A QX ONE ddPCR instrument from BioRad was used to quantify in-frame and out-of-frame dystrophin mRNA (**Figure 1**) and glyceraldehyde 3-phosphate dehydrogenase (GAPDH). The percent of exon skipping was calculated as in-frame / (in-frame + out-of-frame) dystrophin mRNA
- Dystrophin protein was measured by semiquantitative western blot and quantitative IA-UPLC-MS/MS method (**Figure 2**)
 - The western blot method used mouse monoclonal anti-dystrophin antibody MANDYS106 with chemiluminescent detection and myosin heavy chain (MYH) or ACTN2 for normalization
 - The IA-UPLC-MS/MS method used a Sciex Triple Quad 6500+ system and ACTN2 or total protein measurement for normalization

Non-skipped: (out-of-frame)

43 44 51 52 53 54

Skipped: (in-frame)

43 44 52 53 54

The illustration shows a representative genotype of exon 45 to 50 deletion; a skipped ddPCR transcript is only produced when exon 51 is skipped.
 Arrows represent primers, and bars represent fluorescent labeled probes.
 ddPCR, droplet digital PCR

Ab, antibody; ABD, actin-binding domain; CR, cysteine-rich domain; H, hinge region; LLQ, LLQVAVEDR; S, spectrin; IA-UPLC-MS/MS, immunoaffinity ultra-performance liquid chromatography coupled with tandem mass spectrometry; ZF, zinc-finger domain.

Figure 2 reproduced with minor changes from Wang Y, et al. Prenatal diagnosis of Duchenne muscular dystrophy revealed a novel mosaic mutation in dystrophin gene: a case report. *BMC Med Genet.* 2020;11:21(1):222. Licensed under CC BY 4.0 and cropped from original.

- The dystrophin and α -actinin-2 protein concentrations measured by IA-UPLC-MS/MS in 19 healthy adult muscle samples were used as a reference for reporting the percentage of normal dystrophin concentrations in muscle samples from 14 participants with DMD, 1 sample per participant, in study 351-201 (**Figure 3**)
- α -actinin-2/total protein content was correlated with biopsy myofiber content in participants with DMD as measured by hematoxylin and eosin or Masson's trichrome histology (Spearman rank correlation, $P = 0.002$, $R = 0.75$ and $P = 0.02$, $R = 0.067$, respectively)

A.

Muscle trophosphorin (pM)/
total protein (mg/mL)

Participants with

Healthy

DMD, Duchenne muscular dystrophy; IA-UPLC-MS/MS, immunoaffinity ultra-performance liquid chromatography coupled with tandem mass spectrometry.

- Table 1. Head-to-head comparison of western blot vs IA-UPLC-MS/MS human muscle dystrophin assay precision**

*For both detection methods, dystrophin was normalized to α -actinin-2 and the IA-UPLC-MS/MS method was validated to M10. For the comparison, 3 aliquots of each healthy adult muscle sample ($n = 6$) were measured in triplicate (western blot) or duplicate (IA-UPLC-MS/MS) across 3 independent runs.

CV, coefficient of variation; IA-UPLC-MS/MS, immunoaffinity ultra-performance liquid chromatography coupled with tandem mass spectrometry.

Figure 4. Nivudirsen demonstrated dose-dependent exon 51 skipping in all participants

Dose (mg/kg)	Time Point	n	% DMD exon 51 skipping
6	Baseline	6	0.03
	Week 13	2	3.51
9	Baseline	6	0.05
	Week 25	6	5.03

A.

LC-MS dystrophin/g-actinin-2 (% of normal)

Group	Time Point	n	LC-MS dystrophin/g-actinin-2 (% of normal)
6 ma/ka	Baseline	6	0.55
	Week 13	2	2.22
	Week 25	4	2.40
9 ma/ka	Baseline	6	1.16
	Week 25	6	5.01

Results are presented as mean $\pm$ SEM. Week 13 biopsies were from cohort 1A, and week 25 biopsies were from cohorts 1B and 2.

A.

	QC samples				Study samples	
	25%	10%	4%	1%	Baseline	Week 25
Dystrophin						
α-actinin-2						

For panel B, the western blot image revealed multiple dystrophin isoforms with unknown functional capacity at baseline and a higher molecular weight dystrophin isoform after nivolumab treatment.

DMD, Duchenne muscular dystrophy; QC, quality control; W, week.

B.

Baseline	W25

- Figure 7. Exon-skipped dystrophin mRNA correlated with dystrophin protein measured by IA-UPLC-MS/MS after nivudirsen treatment**

Scatter plot showing the correlation between LC-MS dystrophin (pM)/total protein (µg/mL) on the x-axis and Exon 51-skipped DMD/GAPDH mRNA on the y-axis. The plot shows a strong positive correlation with $R = 0.84$ and $P = 0.00016$. Data points are represented by dark blue circles.

The Spearman rank correlation is presented.
DMD, mRNA encoding dystrophin protein; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; IA-UPLC-MS/MS, immunoaffinity ultra-performance liquid chromatography coupled with tandem mass spectrometry.

- Together, these orthogonal methods provide a robust dataset that demonstrates early participant responses and support pharmacodynamic activity consistent with potential clinical benefit in participants treated with nivudirsen
- IA-UPLC-MS/MS and western blot are complimentary methods of protein measurement that offer quantitative and qualitative assessments, respectively
 - Whereas western blot is considered semi-quantitative, IA-UPLC-MS/MS provides absolute quantification of dystrophin based on an internal standard and is less susceptible to sample decay and lab-specific conditions

1. Bladen CL, et al. *Hum Mutat.* 2015;36(4):395–402. 2. Duan D, et al. *Nat Rev Dis Primers.* 2021;7(1):13. 3. Oppeneer T, et al. *Nucleic Acid Ther.* 2025;35(2):68–80. 4. Oppeneer T, et al. *Nucleic Acid Ther.* 2025;35(2):81–92.

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